



TITLE: Effects of bearberry, parsley and corn silk extracts on diuresis, electrolytes composition, antioxidant capacity and histopathological features in mice kidneys

AUTHORS: Marijana Vranješ, Boris M. Popović, , Dubravka Štajner, Vesna Ivetić, Anamarija Mandić, Dejan Vranješ

This article is provided by author(s) and FINS Repository in accordance with publisher policies.

The correct citation is available in the FINS Repository record for this article.

NOTICE: This is the author's version of a work that was accepted for publication in *Journal of Functional Foods*. Changes resulting from the publishing process, such as peer review, editing, corrections, structural formatting, and other quality control mechanisms may not be reflected in this document. Changes may have been made to this work since it was submitted for publication. A definitive version was subsequently published in *Journal of Functional Foods*, Volume 21, March 2016, Pages 272–282. DOI: 10.1016/j.jff.2015.12.016

This item is made available to you under the Creative Commons Attribution-NonCommercial-NoDerivative Works – CC BY-NC-ND 3.0 Serbia



1
2 **Effects of bearberry, parsley and corn silk extracts on diuresis, electrolytes**
3 **composition, antioxidant capacity and histopathological features in mice kidneys**

4
5 **Marijana Vranješ^a, Boris M. Popović^b, Dubravka Štajner^{b1}, Vesna Ivetić^c, Anamarija Mandić^d,**
6 **Dejan Vranješ^a**

7 ^a*Emergency Centre, Clinical Centre of Vojvodina, Hajduk Veljkova 1, 21000 Novi Sad, Serbia*

8 ^b*Faculty of Agriculture, University of Novi Sad, Trg Dositeja Obradovića 8, 21000 Novi Sad, Serbia;*

9 ^c*Faculty of Medicine, University of Novi Sad, Hajduk Veljkova 1, 21000 Novi Sad, Serbia*

10 ^d*Institute for Food Technology, University of Novi Sad, Bulevar cara Lazara 1, 21000 Novi Sad,*

11
12 **A B S T R A C T**

13 The effects of corn silk (*Maydis stigma*), parsley (*Petroselinum crispum*) and bearberry leaves (*Uvae ursi* folium)
14 extracts on diuresis, electrolytes composition, antioxidant capacity and histopathological features of pretreated mice
15 kidneys were determined. The first group of ten animals drank corn silk, second parsley and third bearberry leaf extract.
16 Fourth group was the control when animals drank water. Extracts and water were administrated *ad libitum*. On 0, 1, 7,
17 14 and 28th day of the experiment urine volume and electrolyte content were measured. Antioxidant status of kidneys
18 was investigated by determining antioxidant enzymes and reduced glutathione quantity. Free radical scavenging
19 capacity, lipid peroxidation and total antioxidant power were determined. Histopathological examination of kidneys was
20 performed at the end of the experiment. On the basis of the overall presented results parsley and corn silk extracts could
21 be proposed for further investigations as new functional food ingredients with antioxidant and diuretic properties.

22
23 *Keywords: Maydis stigma, Petroselinum crispum, Uvae ursi, pharmacodynamic-estimation, antioxidant-status,*
24 *histopathological- features*

25

¹ *Corresponding author: Tel:+381 21 485 3374; fax: +381 21 450 857.*
E-mail address. stajnerd@polj.uns.ac.rs (D.Štajner).

26 **1. Introduction**

27 Spices and herbs have been used since antiquity as food supplements to improve flavor of
28 food as well as in traditional medicine in many countries. Nowadays, they have also been the subject
29 of study, particularly by the pharmaceutical and food industries as functional food ingredients
30 because of their potential use for improving health.

31 Antioxidant properties are among the first links between chemical reactions and biological
32 activity and have been extensively studied during the past years. It is generally accepted that free
33 radicals play an important role in the development of oxidative stress and tissue damage and
34 pathological events (Feher et al., 1987; Auroma 1988; Halliwell & Gutteridge, 1989 ; Ahmad & Beg,
35 2013). In recent years, antioxidants from natural food, spices and herbs have attracted increasing interest
36 compared with chemical antioxidants (Zhang et al.,2014; Luthria et al., 2015). Oxidative stress is
37 frequently associated with, and is partly involved in, the pathogenesis of chronic renal failure,
38 hypertension and their complications. Furthermore chronic kidney disease is a significant risk factor
39 for premature cardiovascular disease and death. Increased oxidative stress in people with chronic
40 kidney disease is a significant risk factor for diabetic nephropathy, premature cardiovascular disease
41 and death (Rojas-Rivera et al., 2012).

42 There is an increasing interest in the natural antioxidants in different herbs and their role in
43 health and disease (Shahidi, 1997; Štajner et al., 2006). In addition to the protective effects of the
44 endogenous antioxidant defense system, natural products with antioxidant activity could retard the
45 oxidative damage of a tissue by increasing those defenses (Keli et al., 1996; Schinella et al., 2000 ;
46 Štajner et al., 2014-a; Štajner et al., 2014-b). A diet rich in fruits and vegetables has a positive
47 impact on several chronic conditions, such as obesity, diabetes, cancer, cardiovascular, kidney and
48 neurodegenerative diseases. In some cases fruits as functional foods, are consumed due to their
49 antioxidant properties (Gustavo et al., 2013). Herbal medicines, one of the nature's gifts to

50 mankind, are used for many therapeutic and prophylactic effects, among others, as diuretics. Also,
51 medicinal plants are famous for their antioxidant properties. Herbal diuretics with the longest
52 tradition in folk medicine and modern phytotherapy in Europe, Asia and America include corn silk,
53 parsley and bearberry leaves. Investigation of corn silk, parsley and bearberry leaf extracts effects
54 on diuresis and antioxidant status may point toward new therapeutic approach of different chronic
55 conditions. Spices and other food ingredients used in ethnomedicine may be beneficial to the
56 patients with the kidney disease.

57 Kidney is one of vital organs in the body which removes metabolic waste products, such as
58 urea, creatinine and ammonia. Also kidney keeps a balance of extracellular fluid volume,
59 concentration of inorganic electrolytes in the extracellular fluid, extracellular fluid osmolarity, acid-
60 base balance and blood pressure. In addition, kidney plays a role in producing vitamin D and
61 hormones (National Kidney Federation, 2003). If kidney is damaged, it can no longer perform the
62 function of excretion properly, which leads to accumulation of metabolic waste products. In
63 addition, kidney failure is often followed by a variety of other disorders, such as cardiovascular
64 disease, anemia, osteodystrophy, acidosis, etc. The incident of kidney failure increases in the
65 diabetic and hypertension population (CDC, 2010). Treatments in already advanced kidney failure
66 are mainly symptomatic, and only kidney replacement therapy, such as dialysis or kidney
67 transplantation are treatment of choice in terminal stages. Therefore, safe and non-expensive
68 alternative therapies could be use to prevent the progression of kidney disease (Sukandar et al.,
69 2013).

70 Bearberry, parsley and corn silk are selected as plants which extracts are traditionally used as
71 diuretics. Also, recent works pointed the use of these plants as functional food ingredients
72 (Amarawicz & Pegg, 2013; Jia et al., 2014; Sarepoura et al., 2015). However, their biological
73 properties are actually not completely elucidated (Aw et al., 2014).

74 It is well known that bearberry (*Arctostaphylos uva-ursi* L. Sprengel), parsley (*Petroselinum*
75 *crispum*) and corn silk (*Zea mais* L.hair) are used in treatment and prevention of renal diseases
76 (Amarowicz et al., 2004; Markel, 2005). Folk medicine around the world has recommended *Uva*
77 *ursi* for nephritis, kidney stones, and chronic cystitis. Bearberry extract also known as *Uva ursi*, is
78 considered a potent diuretic so it can enhance the excretion of fluids from the body. Bearberry
79 decreases the accumulation of uric acid, a natural component of urine. Uric acid build up may
80 crystallize and get deposited into the kidneys, joints and blood stream. Bearberry supports the
81 membranes of the urinary system and express antioxidant and antimicrobial activities. Due the high
82 tannin content bearberry-leaf extract can decrease lipid oxidation (Carpenter et al., 2007). *Uva ursi*
83 juice is used to prevent some stomach and kidney problems by suppression of lipid peroxidation and
84 mobilizing of antioxidant activity (Shikov et. al., 2014). Although the use of *Uva-ursi* folium has a
85 historic tradition, controlled clinical studies are needed to verify if indeed *Uva ursi* is effective in
86 humans. It is also important to note that *Uva ursi* can be toxic since hydroquinone can cause serious
87 liver damage. Amarawicz and Pegg (2013) indicated that phenolic compounds extracted from
88 bearberry leaves are potential functional food ingredient with antiproliferative activities, especially
89 against colon carcinoma cell line.

90 *Petroselinum crispum* (parsley) aqueous seed extract was reported to produce a diuretic
91 effect in rats. The mechanism of action of parsley seems to be mediated through an inhibition
92 of the Na^+/K^+ pump that would lead to a reduction in Na^+ and K^+ reabsorption leading to an osmotic
93 water flow into the lumen, and diuresis (Kreydiyyeh & Usta, 2002). Also, *Petroselinum crispum*
94 prevented formation of calcium oxalate stones in rats with nephrolithiasis and reduced the number
95 of calcium oxalate deposits (Saeidi et al., 2012). Jia et al. (2014) proposed parsley as a nutraceutical
96 and functional food intervention in inflammatory bowel disease by multi-omics evaluation.

97 Health benefits of corn silk (*Zea mais* L., hair) have been reported in many investigations.
98 Corn silk extract could promote insulin production in animals, support the recovery of the injured
99 cells of the kidney, pancreas and control blood sugar level in rats (Sarepoua et al., 2013.) Corn silk,
100 a diuretic, can increase the flow of urine from the body and may reduce the risk of new stones
101 developing. It was indicated that corn silk might reduce or even prevent renal damage by defending
102 kidney against oxidative stress (Sukandar et al., 2013). Silk of corn, rich in polyphenol compounds
103 can be used as dietary fiber and as a food additive for the prevention of several diseases (Sarepoua et
104 al., 2015).

105 The aim of this study was to investigate corn silk (*Maydis stigma*), parsley (*Petroselinum*
106 *crispum*) and bearberry leaf (*Uvae ursi* folium) extracts as potential functional food ingredients
107 with the effects on diuresis and electrolyte composition of urine and to evaluate the effects of these
108 extracts on antioxidant capacity of kidneys. Furthermore the effect of *Maydis stigma*, *Petroselinum*
109 *crispum* and *Uvae ursi* folium extracts on histopathological features of the kidneys of pretreated
110 experimental animals was also examined.

111

112 **2. Materials and methods**

113 *2.1. Chemicals and reagents*

114 Chemicals used in the experiments: 2,2'-diphenyl-1-picrylhydrazyl (DPPH), 2,4,6-tripyridyl-
115 *S*-triazine (TPTZ), nitro blue tetrazolium (NBT), quaiacol (2-methoxyphenol), Folin & Ciocalteu-
116 phenol reagent, 2-thiobarbituric acid (TBA), ferric chloride and monobasic potassium phosphate
117 were obtained from Sigma-Aldrich (Beograd, Serbia). Methanol (HPLC, gradient grade), quercetin
118 and formic acid were supplied by Merck KGaA (Darmstadt, Germany). Standard substances
119 including gallic acid, protocatechuic acid, catechin, caffeic acid, vanillic acid, chlorogenic acid,
120 ferulic acid, rutin, quercetin, myricetin, luteolin, kaempferol, apigenin and orientin were purchased

121 from Sigma-Aldrich GmbH (Sternheim, Germany). Water used throughout the experiments was
122 purified using a Millipore, Elix UV and Simplicity Water Purification System (Milford, MA, USA).

123 2.2. *Experimental animals*

124 The study was conducted on experimental animals body weight of 31-46 g and ages up to 3
125 months (*Mus musculus* variety albino NMRI), which were selected randomly from the litter of
126 Pasteur Institute in Novi Sad, Serbia. Research was conducted in accordance to the principles
127 established for research in animal models (European Council Directive of November 24, 1986,
128 86/609/EEC, III-2014-04). Animal care and all experimental procedures were conducted in
129 accordance with the Guide for the Care and Use of Laboratory Animal Resources, edited by
130 Commission of Life Sciences, National Research Council, Male and Female Hanover National
131 Medical Institute (Hann NMRI). Mice were bred in the vivarium at the Department of
132 Pharmacology, Toxicology and Clinical Pharmacology, Medical Faculty, University of Novi Sad,
133 Serbia. Animals were kept in the metabolic plexiglas cages at constant room temperature $21 \pm 1^\circ\text{C}$
134 and humidity $55\% \pm 1.5\%$, with circadian rhythm (day/night). They were fed with the standard
135 laboratory mice feed, produced by the Veterinary Institute in Zemun, Serbia. Animals were given
136 free access to food and fluid (water or plants extracts). Experimental animals were divided into four
137 groups of ten animals. During the experiment the first group of ten animals drank corn silk extract,
138 the second group parsley extract, the third group bearberry leaf extract *ad libitum*. The fourth group
139 was control group in which animals drank water *ad libitum*. Urine of all groups was collected on 0,
140 1, 7, 14 and 28h day of experiment, and on the same date urine volume was measured as well as
141 electrolyte contents. All animals were sacrificed under urethane anesthesia. Kidneys were removed
142 and homogenized and after extraction, antioxidant status was determined using different
143 contemporary methods.

144 2.3. *Herbal material extraction*

145 Herbal material which was used in this study was of controlled origin (mountain Rtanj,
146 2014) available in retail. The plant leaves were dried in air and ground in a mixer. An amount of 200
147 g of the finely powdered material (0.2-0.3 mm) was extracted with 4 L of 96% ethanol (EtOH),
148 during 72h at room temperature (25°C). After that, extracts were vacuum evaporated at 40°C. Dry
149 residues were used to prepare 5% aqueous extracts used in the experiment for polyphenol
150 characterization and total phenol determination.

151

152 *2.4. Biochemical Assays*

153 *2.4.1. Serum urine analyses*

154 Concentrations of blood urea nitrogen and creatinine were estimated as described by
155 Fossati et al. (1980), respectively using specific diagnostic kits (Sigma Aldrich, St. Louis, MO,
156 USA). Serum and urine levels of sodium and potassium were determined as described by Ali (2010).

157

158 *2.4.2. Preparation of kidney homogenate*

159 One gram of the right kidney tissue was collected, washed in ice-cooled 0.9% NaCl and
160 homogenized in ice-cooled 1.15% potassium chloride solution and 50 mM potassium phosphate
161 buffer solution (pH 7.4) to yield 10% homogenate (w/v). Homogenization was performed using
162 ultrasonic homogenizer. The homogenate was then centrifuged at 4000 x g for 5 min at 4°C. The
163 supernatant was collected and kept for further use.

164

165 *2. 5. Assessment of prooxidant / antioxidant activity*

166 Prepared kidney homogenate was used for antioxidant determinations (DPPH RSC, FRAP
167 and ·OH quantity) determination as well as enzymatic assays, glutathione and lipid peroxidation.

168 The superoxide dismutase, SOD activity was determined in aliquots by the method of Misra
169 and Fridovics (1972), based on the inhibition of transformation of adrenaline to adrenochrome at
170 pH 10.2 (Matkovics et al., 1977) ; guaiacol peroxidase GPX activity, using guaiacol as substrate
171 (Matkovics et al., 1977).; glutathione peroxidase GSH-Px activity using cumene hydroperoxide and
172 reduced glutathione (GSH) as substrates (Chiu et al., 1976); catalase CAT activity
173 spectrophotometrically at 240 nm (Beers et al., 1952; Iwase et al., 2013); Lipid peroxidation was
174 measured by TBARS (quantities of thiobarbituric acid reactive substances) assay and values were
175 reported as equivalents of malonyldialdehyde (MDA); the calibration curve was prepared with
176 malonyldialdehyde bis-diacetal (Placer et al., 1968); hydroxyl radical by the inhibition of
177 deoxyribose degradation and expressed by nmol OH• in the model system (Cheesman et al., 1988).

178 The quantity of GSH was determined with Ellman reagent (Sedlak & Lindsay, 1968) and
179 protein content according to Bradford (1976).

180 Radical scavenging capacity (RSC) was determined using 1, 1-diphenyl-2-picrylhydrazyl
181 radical (DPPH•). Reduction of DPPH radical was determined measuring disappearance of DPPH at
182 515 nm. RSC is expressed by percents compared to the control (Abe & Hirota, 1998). The percent
183 inhibition of the DPPH radical (RSC) by the samples was calculated using the formula:

$$184 \quad \text{RSC} = [(A_c - A_x) / A_c] \times 100 \%$$

185 where A_c is absorbance of the control and A_x is absorbance of the sample after 30 min of incubation.

186 Total antioxidant capacity was estimated according to the FRAP (Ferric Reducing
187 Antioxidant Power) assay (Benzie & Strain, 1999). Total reducing power is expressed as FRAP
188 units. FRAP unit is equal with $100 \mu\text{mol}/\text{dm}^3 \text{Fe}^{2+}$. FRAP value was calculated using formula:

$$189 \quad \text{FRAP value} = \Delta A_{\text{sample}} / \Delta A_{\text{standard}}$$

190

191 *2.6. Polyphenol determination in plant extracts*

192 The total phenolic content was determined by a modification of the Folin-Ciocalteu method
193 and the results expressed as mg gallic acid/g extract (Vinson et al., 2003).

194 Ethanolic extracts of the investigated plants (bearberry, parsley and corn silk leaves) were
195 used for HPLC analysis. HPLC analysis was performed by using a liquid chromatograph (Agilent
196 1200 series; Agilent Technologies, Palo Alto, CA, USA), equipped with a diode array detector
197 (DAD), Chemstation Software (Agilent Technologies), a binary pump, an online vacuum degasser,
198 an autosampler and a thermostated column compartment, on an Agilent, Eclipse XDB-C18, 1.8 μm ,
199 4.6 \times 50 mm column, at a flow-rate of 1 mL min⁻¹. Solvent gradient was performed by varying the
200 proportion of solvent A (methanol) to solvent B (1% formic acid in water (v/v)) as follows: initial
201 10% A; 0-10 min, 10 -25% A; 10-20 min, 25 - 60% A; 20-30 min, 60-70% A. The total running
202 time and post-running time were 45 and 10 min, respectively. The column temperature was 30°C.
203 The injected volume of samples and standards was 5 μL and it was done automatically using
204 autosampler. The spectra were acquired in the range 210–400 nm and chromatograms plotted at 280,
205 330 and 350 nm with a bandwidth of 4 nm, and with reference wavelength/ bandwidth of 500/100
206 nm. Content of the investigated plant phenolic compounds in crude extracts is expressed as mg g⁻¹
207 extract (Mišan et al., 2011).

208 *2.7. Histological procedure*

209 Kidney specimens were taken and fixed in 10 % neutral formalin solution. The fixed
210 specimens were dehydrated in ascending grades of alcohol, cleared in xylene. They were embedded
211 in paraffin boxes, sectioned at 4-6 microns thickness, stained with Hematoxylen and Eosin (H&E)
212 and examined microscopically according to Carleton (1976).

213

214 2.8. *Statistical evaluation*

215 All determinations were performed in triplicate and 10 experimental animals. Data were
216 expressed as mean \pm standard error (SE). Values were means for ten mice. Statistical comparisons
217 between samples were performed with Duncan t-test (Snedecor & Cochran, 1986), for independent
218 observations were done using STATISTICA 9.1. Differences were considered significant at $p <$
219 0.05.

220

221 **3. Results and discussion**

222 The kidneys function is to excrete toxins through the urine and balance out our electrolytes
223 (sodium, potassium, calcium, magnesium, chloride and phosphorous) and amino acid levels. It is
224 especially important for effective elimination of ammonia, which is highly toxic and destructive to
225 brain tissue.

226

227 *3.1. Pharmacodynamic investigations*

228

229

230

231 Table 1- presents the results concerning urine pharmacodynamic investigations. The effects
232 of 5% water extracts of *Maydis stigma*, *Petroselinum crispum* and *Uvae ursi folium* on diuresis, K^+
233 and Na^+ quantities.

234

Table 1.

235 On the basis of pharmacodynamic results presented in Table 1, it is possible to observe that
236 the best diuretic effect was achieved on the second day of experiment under parsley influence. It is
237 well known that parsley has high antioxidant ability and also provokes an osmotic water flow into
238 the lumen, and diuresis (Kreydiyyeh & Usta, 2002). The corn silk extract induced maximum diuresis
239 on the first day of the experiment. The lowest diuretic effect was observed under bearberry influence
where maximum was also reached on the first day of experiment (Table1).

240 Comparing the results from all three experimental groups after seven days of administration,
241 parsley extract induced the strongest diuretic effect. The best effect on K^+ quantity in urine was
242 observed under the action of parsley and corn silk. After seventh day of experiment, the decrease in
243 K^+ quantity was observed under the action of parsley and corn silk extracts. Bearberry extract
244 induced K^+ quantity increase effect at the first day of administration but smaller comparing to the
245 corn silk and parsley. After seventh days of administration, K^+ quantity induced by bearberry was
246 under the control values. Na^+ quantity reached a maximum at seventh day (parsley extract) or first
247 two days (corn silk). The lowest Na^+ quantity was observed under bearberry action at the first day of
248 administration. After the first day of bearberry extracts administration significant decrease of Na^+
249 quantity comparing to the control group was observed (Table1).

250 The nephroprotective effect of *Petroselinum crispum* herb, reported in the present study, was
251 similar to that reported by Afzal et al., 2004, who found that a polyherbal formulation containing
252 *Petroselinum crispum* (parsley) produced a nephroprotective effect in mice. This effect of
253 *Petroselinum crispum* was attributed to its *in vitro* antioxidant activity as free radical scavenger or
254 due to its high content of flavonoids (Fejes et al., 1998). The diuretic effect of *Petroselinum*
255 *crispum* was reported by Kreydiyyeh and Usta (2002), who found that parsley aqueous seeds extract
256 produced a diuretic effect in rats. The previous authors concluded that the mechanism of action of
257 parsley seems to be mediated through an inhibition of the Na^+/K^+ pump that would lead to a
258 reduction in Na^+ and K^+ reabsorption thus leading to an osmotic water flow into the lumen, and
259 diuresis. Urinary potassium and sodium levels could be changed as the consequence of longer
260 administration of different agents (Ali, 2010), as it happen also under influence of *Maydis stigma*
261 *and Petroselinum crispum* extracts during seven days of administration.

262 In Table 2, the results concerning the effects of extracts of *Maydis stigma*, *Petroselinum*
263 *crispum* and *Uvae ursi* folium on BUN (blood urea nitrogen) and serum creatinine are presented.

264 Comparing control values, the best effect in urea and creatinine reduction was observed under the
265 extracts of corn silk and parsley. Bearberry extract reduced urea content comparing to the control,
266 but slightly increased the content of creatinine.

267 Table 2.

268
269 Results presented in Table 2. Supported the results from Table 1. They connected and
270 explained pharmacodynamic effects, with urea and creatinine reduction under the influences of
271 *Maydis stigma* and *Petroselinum crispum* extracts.

272
273 *3.2. Prooxidant / antioxidant activity*

274 The effects of bearberry, parsley and corn silk extracts, on SOD, CAT, GPx and GSH-Px
275 activities in mice kidneys are presented in Fig. 1. SOD activity was higher in kidneys of mice treated
276 with corn silk extract (16.03 U/mg protein) and lowest in kidneys of mice treated with parsley
277 extract (12.33 U/mg protein). In kidneys of animals which drank bearberry extract (14,05 U/mg
278 protein) SOD was slightly smaller from the control value (14.05 U/mg protein). Statistically
279 significant differences concerning the control value were observed under parsley and corn silk
280 influence. Bearberry extract did not provoke significant effect. Our results implicated that corn silk
281 provoked the most favorable effect to SOD activity what is in agreement with results of other
282 authors (Hua et al., 2010), which proved that *Maydis stigma* had antioxidative and protective role in
283 kidneys and liver of experimental animals (Fig 1A).

284 Fig 1B. presents the effects of bearberry, parsley and corn silk extracts on CAT activity of
285 mice kidneys . CAT activity was high in kidneys of animals that consumed parsley extract (67.21
286 nmol H₂O₂/mg protein) and in the control sample (66.72 nmol H₂O₂/mg protein) but lowest in
287 kidneys of animals drank with bearberry extract, 46.85 U/mg protein. Kidneys of animal which
288 drank corn silk extract, exhibited low CAT activity of 49.11 nmol H₂O₂/mg protein. (Fig.1B.).

289 Statistically significant differences concerning control value were observed under bearberry and
290 corn silk influence, though parsley did not provoke significant effect.

291 Fig. 1

292 Presented results (Fig.1A. and Fig 1B.) are in the agreement with results that mice kidneys exhibited
293 substantial activities of antioxidant enzymes (Lenzen et al., 1996).

295 The effects of bearberry, parsley and corn silk extracts, on GPx activity of mice kidneys are
296 presented in Fig. 1C. GP_x activity was higher in kidneys of animals which drank bearberry extract
297 (151,01 nmol guajacol/mg protein). GPx activities were substantially lower in kidneys of animal
298 which drank the extracts of other two medicinal plants. The lowest was in kidneys of mice treated
299 with parsley extract, 48.58 nmol guajacol/mg protein. Statistically significant differences concerning
300 the control value were observed under the influence of all used plants extracts. (Fig 1C.). High GPx
301 in kidneys and other mice organs play important role in defense from different inflammatory
302 processes (Duget et al., 2001).

303 GSH-Px activity was high in kidneys of animals which drank corn silk (75.18 nmol GSH/mg
304 protein) and bearberry (69.85 nmol GSH/mg protein) extracts as presented in Fig.1D. Control
305 sample exhibited the smallest GSH-Px activity of 28.93 nmol GSH/mg protein. Statistically
306 significant differences concerning the control value occurred under the treatments with bearberry
307 and corn silk. Our results pointed to high GSH-Px activity in kidneys, what is in connection with
308 enzymic functioning of detoxification (Klivenyi et al., 2000). It was also proved that the use of
309 plants extracts increased activities of antioxidant enzymes in mice (Caia & Weib 1996), and
310 therefore improved abilities to reduce the consequences induced by different types of oxidative
311 stress (Güven & Gülmez 2003), what is in agreement with our results presented in Fig. 1.

312 Effects of *Maydis stigma*, *Petroselinum crispum* and *Uvae ursi* folium extracts, on reduced
313 glutathione quantity, ferric reducing antioxidant power and soluble protein content in mice kidneys
314 are presented in Fig.2.

315 Fig.2.

316 Quantity of GSH in mice kidneys was affected by the action of all examined medicinal
317 plants Fig. 2A. GSH quantity was high in kidneys of animals which drank corn silk extract (187.14
318 nmol/mg protein) and in the control sample (179.50 nmol/mg protein) and lower in kidneys of
319 animals treated with parsley extract, 143.02 nmol/mg protein. Statistically significant differences
320 concerning the control value occurred under treatment with corn silk. It was proved that high
321 quantities of GSH in kidneys are connected with induced tolerance of mice to oxidative stress (Saito
322 et al., 2010). Oral administration of *Petroselinum crispum* expressed nephroprotective and diuretic
323 effects, as it reversed the biochemical and antioxidant activity as evident by decreasing lipid
324 peroxidation, increasing content of reduced glutathione and restoring activities of antioxidant
325 (SOD, GPx and CAT) enzymes in renal tissue (Shalby et al., 2014).

326 Considerable influence of parsley and corn silk extracts on FRAP values in kidneys of the
327 experimental animals was observed (Fig. 2B). Bearberry extract induced the highest FRAP value
328 (24.17 $\mu\text{mol Fe}^{2+}/\text{g}$), and parsley extract, the lowest 20.51 $\mu\text{mol Fe}^{2+}/\text{g}$. All used medicinal plants
329 increased FRAP values comparing to the control (7.33 $\mu\text{mol Fe}^{2+}/\text{g}$). Statistically significant changes
330 were observed under the influence of all three used medicinal plants. Due to our results presented in
331 Fig. 2B., it was obvious that all FRAP values in kidneys of mice increased under influence of
332 investigated medicinal plants. Similar effects induced antioxidants from red vines (Rodrigo et al.,
333 2002).

334 Results concerning the effect of bearberry, parsley and corn silk extracts on soluble proteins
335 content in kidneys of experimental animals were shown in the Fig.2C. The content of soluble

336 proteins was pretty equal, but still the highest under the influence of parsley extract (62.82 mg/g).
337 Significant changes were observed only under parsley influence.

338 The effects of bearberry, parsley and corn silk extracts on DPPH radical scavenging capacity
339 (RSC) in kidneys of mice are presented in Table 3. DPPH RSC was improved by the action of all
340 three medicinal plants. IC₅₀ values were lower in samples treated with medicinal plants than in the
341 control sample (4.64 mg/cm³). The highest RSC was observed in kidneys of mice treated with
342 parsley extract (2.62 mg/cm³) and lowest in kidneys of mice treated with corn silk extracts (3.48
343 mg/cm³). DPPH scavenging is often used to assess the antioxidative effects of different plants in
344 tissues of experimental animals in order to prevent different diseases (Alvares- Gonzales et al.,
345 2014).

346 Table 3.

347 The effects of bearberry, parsley and corn silk extracts on •OH and lipid peroxidation are
348 presented in Fig.3. •OH quantity in kidneys changed under the influence of medicinal plants
349 (Fig.3A). It was the highest in control sample (3.10 U nmol/mg protein), but in samples treated with
350 medicinal plants it was almost at the same lower level. Statistically significant changes were
351 observed under the influence of bearberry and parsley extracts. Control sample accumulated higher
352 quantity of •OH than after the treatment with medicinal plants what proves that they diminished
353 •OH generation and acted protectively (Li et al., 1999; Ueno et al., 2007). Statistically significant
354 changes were observed under the influences of all investigated medicinal plants.

355 Fig.3.

356 Bearberry, parsley and corn silk influenced LP in kidneys of experimental animals (Fig 3B).
357 Lipid peroxidation was reduced under the influence of all three medicinal plants and ranged from
358 3.64 nmol MDA equivalents/mg protein in kidneys of animals which drank bearberry extract to 4.20
359 nmol MDA equivalents/mg protein in kidneys of animals which drank corn silk extract. The highest

360 LP was observed in the control group of animals, 4.80 nmol MDA equivalents/mg protein.
361 Statistically significant changes were observed under the influences of all investigated medicinal
362 plants. Lipids containing polyunsaturated fatty acids are readily oxidized by molecular oxygen and
363 such oxidation proceeds by a free radical chain mechanism causing tissue damages (De Groot, &
364 Noll, 1987). Oral administration of *Petroselinum crispum* expressed nephroprotective and diuretic
365 effects, as it reversed the biochemical and antioxidant activity as evident by decreasing lipid
366 peroxidation, increasing content of reduced glutathione and restoring activities of antioxidant
367 (SOD, GPx and CAT) enzymes in renal tissue (Shalaby et al., 2014). Other authors suggested that
368 corn silk extract induces reduction of LP and increase of catalase and SOD activities and therefore
369 could be used in repairing renal damages (Sukandar et. al.,2013) what is in agreement with our
370 results presented in Figures 1-3. Antioxidants also significantly controlled the speed of lipid
371 peroxidation and the reduction of the activities of SOD, GSH-Px, and CAT (Wu et al., 2013).
372 Similar effects were observed in kidney tissues under influence of *Maydis stigma* and *Petroselinum*
373 *crispum*.

374 3.3. Polyphenol characterization of plant extracts

375 In Table 4, the results concerning polyphenol constitution of bearberry, parsley and corn silk
376 extracts are presented.

377 Table 4.

378 Total content of phenol compounds determined by Folin-Ciocaltey method was: 95 mg/g bearberry
379 extract; 119 mg/g parsley extract and 110 mg/g corn silk extract. Sum of phenolics determined by
380 HPLC was 34.8 mg/g bearberry extract; 10.39 mg/g parsley extract and 4.21 mg/g corn silk extract.
381 Regardless to the highest content of phenolic compounds found in bearberry by HPLC, parsley
382 extract showed the greatest diversity of present phenolic structures including different phenolic
383 acids, but the dominant component of parsley extract were kaempferol and kaempferol derivatives

384 (5.84 mg/g). Dominant compounds in bearberry extract were rutin and its derivatives (13.74mg/g),
385 catechin and its derivatives (12.23 mg/g) and gallic acid (7.75 mg/g). The dominant phenolic
386 compounds in corn silk were orientin derivatives (3.24 mg/g). Presented results are in the accordance
387 with previous polyphenol characterization of bearberry, parsley and corn silk extracts (Panusa et al.,
388 2015; Yidiz et al., 2015 and Liu et al., 2012). The best diuretic effect, the effect on electrolyte
389 excretion and DPPH antiradical activity showed parsley extract which could be attributed to the
390 presence of wide range of phenolic compounds, especially kaempferol and its derivatives. As
391 phenolic compounds are classical antioxidant agents, this effect could increase the bioavailability of
392 endothelial nitric oxide leading to dilatation of the renal afferent arterioles and increasing glomerular
393 filtration rate (Gasparotto Junior et al., 2012). Although corn silk contained the poorest profile of
394 polyphenols in crude extract, it showed significant positive effect on SOD and GSH-Px activity in
395 mice kidneys, probably because of the orientin derivatives presence. Bearberry extract, full of rutin
396 and catechin derivatives could be probable cause of peroxidase induction in mice kidneys. These
397 initial observations and findings suggest that the ethanolic extracts of parsley, bearberry and corn
398 silk leaves contain different chemical compounds which biological potential activity deserves
399 further investigation.

400

401 3.4. *Histopathological examinations*

402 Histological examination of kidneys of healthy mice showed normal histological structure of
403 renal parenchyma (glomeruli and tubules) as illustrated in Fig. 4A.

404 Fig.4.

405
406 Histopathological studies showed that bearberry extract induced significant changes on
407 kidney parenchyma (Figs 4B. and 4C.). *Uvae ursi* folium extract caused microcystic changes with

408 visible protein hyalin cylinder, focal bleeding, peritubular exudation and pertubular nephritis
409 especially in the regions with cystic changes. Under influence of *Maydis stigma* and *Petroselinum*
410 *crispum* extracts, no harmful changes were observed comparing to the control kidney tissue.

411 412 4. *Conclusion*

413
414 The results obtained in this work demonstrated that all three tested plants differently
415 influenced physiological function of kidney, as well as their antioxidant capacity and
416 histopathological features. The best diuretic effect which was maintained for the longest time, was
417 observed under the influence of parsley extract full of polyphenols including phenolic acids and
418 flavonols. Furthermore, parsley extract showed the best results concerning electrolyte excretion.
419 Most pronounced reduction in BUN (blood urea nitrogen) and creatinine was observed by the action
420 of corn silk extract, and then parsley extract. Overall, high antioxidant capacity was observed in
421 kidneys by the action of all examined medicinal plants, which indicated their high antioxidant
422 capacity. Histopathological examination pointed that kidney parenchyma was significantly
423 influenced by the action of bearberry extract. Other extracts did not change kidney tissue. On the
424 basis of the overall presented results (diuretic, excretion of Na⁺, K⁺, antioxidant, histopathological
425 investigations), parsley extract can be proposed for further investigations as a new therapeutic
426 approach for functional nutraceuticals to be used in the prevention and treatment of kidney disease.
427 The results reported the principle pre-clinical data for application of *Petroselinum crispum* and
428 *Maydis stigma* to attenuate kidney diseases. That also means that they could have huge potential in
429 preventing various kidneys disorders immediately as it appears. As such, further hypothesis testing
430 and mechanistic studies are warranted.

431 *Conflict of interest*

432 None declared.

433

434 **Acknowledgement**

435

436 This research is a part of IPA Project Plantrain (HUSBR/1203/221/173) and the project No. TR
437 31029 which is financially supported by the Ministry of Science, Technologies and Development of
438 the Republic of Serbia.

439

440

441 **REFERENCES**

442 Abe, N., Murata, T., & Hirota, A. (1998). 1,1-diphenyl-2-picrylhydrazyl-radical scavengers,
443 bisorbicillin and demethyltrichodimerol, from a fungus. *Bioscience Biotechnology and Biochemistry*
444 ,62, 661-662.

445

446 Afzal , M., Khan, N.A., Ghufran, A., Iqbal. A. (2004). Inamuddin M. Diuretic and nephroprotective
447 effect of Jawarish Zarooni Sada, a polyherbal formulation. *Journal of Ethnopharmacology* , 91, 219-
448 223.

449

450 Ahmad, S.,& Beg,Z.H.(2013).Alleviation of plasma, erythrocyte and liver lipidemic-oxidative stress by
451 thymoquinone and limonene in atherogenic suspension fed rats. *Journal of Functional Foods*,5, 251–
452 259

453

454 Ali, A.A. (2010). The Effect of long term use of glibenclamide on serum and urinary sodium and
455 potassium levels in Type 2 DM patients. *Iraqi Journal of Pharmaceutical Science*,19: 58-61.

456

457 Álvarez-González,I., Fernando Garcia-Melo, F., Verónica R. Vásquez-Garzón, V., Saúl Villa-
458 Treviño,A. E., Madrigal-Santillán, O., José A. Morales-González, J.A., Jorge A. Mendoza-
459 Pérez,J.A., & Madrigal-Bujaidar, E.. (2014). Evaluation of Blueberry Juice in Mouse
460 Azoxymethane-Induced Aberrant Crypts and Oxidative Damage. *Evidence-Based Complementary*
461 *and Alternative Medicine*, Volume 2014, Article ID 379890, 8 pages.

462

463 Amarowicz, R., R.B. Pegg, R.B., Rahimi-Moghaddam, P., Barl,B. & Weil, J.A. (2004).Free-radical
464 scavenging capacity and antioxidant activity of selected plant species from the Canadian prairies.
465 *Food Chemistry*, 84(4), 551-562.

466

467 Amarowicz, R., R.B. Pegg, R.B. (2013) Inhibition of proliferation of human carcinoma cell lines by
468 phenolic compounds from a bearberry-leaf crude extract and its fractions, *Journal of Functional*
469 *Foods* 5, 660-667.

470

471 Aruoma, O. I. (1998) Free radicals, oxidative stress, and antioxidants in human health and disease.
472 *Journal of the American Oil Chemists' Society*.,75(2),199–212 .

473

474 Aw, H.,J.,W., Hanate, M. Takahashi, S., Saito, K., Tanaka, H., Tomita, M. & Hisanori Kato, M.
475 (2014) Multi-faceted integrated omics analysis revealed parsley (*Petroselinum crispum*) as a novel
476 dietary intervention in dextran sodium sulphate induced colitic mice. *Journal of Functional*
477 *foods*,11, 438-448.

478

479 Beers, R.F. J.r, & Sizer, I.W. (1952) A spectrophotometric method for measuring the breakdown
480 of hydrogen peroxide by catalase. *Journal of Biological Chemistry*, 195(1), 133-40.
481

482 Benzie, I.F.F., & Strain, J. J. (1999). Ferric reducing antioxidant power assay: Direct measure of
483 total antioxidant activity of biological fluids and modified version for simultaneous measurement of
484 total antioxidant power and ascorbic acid concentration. *Methods in Enzymology*, 299, 15-27.
485

486 Bradford, M.M. (1976). A rapid and sensitive method for the quantitation of microgram quantities
487 of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*, 72, 248-253.
488

489 Caia, Q., & Weib, H. (1996). Effect of dietary genistein on antioxidant enzyme activities in
490 SENCAR mice. *Nutrition and Cancer*, 25(1), 1-7.
491

492 Carpenter, R., O'Grady, M.N., O'Callaghan, Y.C., O'Brien, N.M., & Kerry, J.P. (2007). Evaluation
493 of the antioxidant potential of grape seed and bearberry extracts in raw and cooked pork. *Meat
494 Science*, 76, 604–610.
495

496 CDC, 2010. National chronic kidney disease fact sheet 2010.
497 http://www.cdc.gov/diabetes/pubs/pdf/kidney_Factsheet.pdf
498

499 Carleton, H. (1976). *Carleton's Histological technique*. The 4th Edition, (pp. 91-95). London, New
500 York, Toronto: Oxford University Press.
501

502 Cheesman, K.H., Beavis, A., & Esterbauer, H. (1988). Hydroxyl radical induced iron catalyzed
503 degradation of 2-deoxyribose. *Biochemical Journal*, 232, 649-653.
504

505 Chiu, D.T.Y., Stults, F.H., & Tappel, A.L. (1976). Purification and properties of rat lung soluble
506 glutathione peroxidase. *Biochimica Biophysica Acta*, 445, 558-606.
507

508 De Groot, H., & Noll, T. (1987). The role of physiological oxygen partial pressures in lipid
509 peroxidation. Theoretical considerations and experimental evidence. *Chemistry and Physics of Lipids*,
510 44, 209–26.
511

512 Duguet, A., Iijima, H., Eum, S.Y., Hamid, Q., & Eidelman, D.H. (2001). Eosinophil peroxidase
513 mediates protein nitration in allergic airway inflammation in mice. *American Journal of Respiratory
514 and Critical Care Medicine*, 164 (7), 1119-1126.
515

516 Fehér J, Csomós G, Vereckei A. (1987). *Free radical reactions in medicine* (pp. 40–43). Berlin-
517 Heidelberg: Springer-Verlag.
518

519 Fejes, S., Kery, A., Blazovics, A., Lugasi, A., & Lemberkovics, E. (1998) Investigation of the in
520 vitro antioxidant effect of *Petroselinum crispum*. *Acta Pharmacologica Hungarica*, 68(3), 150-156.
521

522 Fossati, P., Principe, L., & G. Berti, G. (1980). Enzymatic colorimetric method for determination of
523 uric acid in serum. *Clinical Chemistry*, 26, 227-237.
524

525 Gasparotto Junior, A., Prando, T.B.L., Leme, T.S.V., Gasparotto, F.M., Lourenço, E.L.B.,
526 Rattmann, Y.D., Silva-Santos, J.E., Kassuya, C.A.L., Marques, M.C.A., (2012). Mechanisms
527 underlying the diuretic effects of *Tropaeolum majus* L. extracts and its main component
528 isoquercitrin. *Journal of Ethnopharmacology* 141, 501-509
529

530 Gustavo, A., Costa, V., Diego, A., Garcia-Diaz, F. , Jimenez, P., Silva, P.H. (2013). Bioactive
531 compounds and health benefits of exotic tropical red–black berries. *Journal of functional foods*,5 ,
532 539-549.
533

534 Güven, A., & Gülmez, M. (2003). The effect of kefir on the activities of GSH-Px, GST, CAT, GSH
535 and LPO levels in carbon tetrachloride-induced mice tissues. *Journal of Veterinary Medicine, Series*
536 *B*, 50(8), 412–416.
537

538 Halliwell, B., & Gutteridge, J.M.C. (1989). *Free Radicals in Biology and Medicine* (pp.416–494).
539 Oxford: Clarendon Press.
540

541 Hua, B., Chunxu, H., Miaomiao, X., Xin, L., & Rui, L. (2010). Protective Effect of Maize Silks
542 (*Maydis stigma*) Ethanol Extract on Radiation-Induced Oxidative Stress in Mice. *Plant Foods for*
543 *Human Nutrition*, 65(3), 271-276.
544

545 Iwase, T., Tajima, I., Sugimoto, S., Ken-ichi Okuda, K., ppei Hironaka, I., Yuko Kamata, Y.,
546 Takada, K., & Mizunoe Y. (2013). A Simple Assay for Measuring Catalase Activity: A Visual
547 Approach. *Scientific Reports*, 3, Article number: 3081 doi:10.1038/srep03081.
548

549 Jia, H., Aw, W., Hanate M., Takahashi, S., Saito, K., Tanaka, H., Tomita M & Kato, H. (2014)
550 Multi-faceted integrated omics analysis revealed parsley (*Petroselinum crispum*) as a novel dietary
551 intervention in dextran sodium sulphate induced colitic mice, *Journal of Functional Foods* 11, 438-
552 448.
553

554 Luthria , D.L., Lu, Y. & John, K.M. (2015) Bioactive phytochemicals in wheat: Extraction,
555 analysis, processing, and functional properties, *Journal of Functional Foods*, in press.
556

557 National Kidney Federation, 2003. End stage renal disease media fact pack.
558 <http://www.kidney.org.uk/campaigns/Press-Media/Press-Guide.pdf>.
559

560 Keli, S.O., Hertog, M.G.L., Feskens, E.J.M., & Kromhout , D. (1996). Dietary flavonoids,
561 antioxidant vitamins, and incidence of stroke: the Zutphen study. *Archives of Internal*
562 *Medicine*,156,637–42.
563

564 Klivenyi, P., Andreassen, O.A., Ferrante, R.J., Dedeoglu N., Mueller, G., Lancelot, E., Bogdanov,
565 M.K., Andersen, J.K., Jiang, D., & Beal, M.F. (2000).Mice Deficient in Cellular Glutathione
566 Peroxidase Show Increased Vulnerability to Malonate, 3-Nitropropionic Acid, and
567 1-Methyl-4-Phenyl-1,2,5,6-Tetrahydropyridine. *The Journal of Neuroscience*, 20(1), 1-7.
568

569 Kreydiyyeh, S.I. & Usta, J. (2002). Diuretic effect and mechanism of action of parsley. *J*
570 *Ethnopharmacol*, 79(3),353-7.
571
572 Lenzen, S., Drinkgern, J., & Tiedge, M. (1996). Low antioxidant enzyme gene expression in
573 pancreatic islets compared with various other mouse tissues. *Free Radical Biology and Medicine*,
574 20(3),463–466.
575
576 Li, B., Gutierrez, P.L., Amstad, P., & Blough, N.V. (1999). Hydroxyl radical production by mouse
577 epidermal cell lines in the presence of quinone anti-cancer compounds. *Chemical Research in*
578 *Toxicology*, 12(11), 1042-1049.
579
580 Liu, J., Wang, C., Wang, Z., Zhang, C., Lu, S. & Liu, J., (2011) The antioxidant and free-radical
581 scavenging activities of extract and fractions from corn silk (*Zea mays* L.) and related flavone
582 glycosides, *Food Chemistry* 126, 261–269.
583
584 Matkovics, B., Novak, R., Hanh Huang, D.U.C., Szabo.I., Varga. Sz.I., & Zelesna, A. (1977). A
585 comparative study on some more important animal peroxide metabolism enzymes. *Comparative*
586 *Biochemistry and Physiology*, 56B, 31-34.
587
588 Misra, H.P., & Fridovics, I.J. (1972). The role of superoxide anion in the autooxidation of
589 epinephrine and a simple measurement for superoxide dismutase. *Journal of Biological Chemistry*,
590 247, 3170-3175.
591
592 Mišan, A., Mimica-Dukić, N., Mandić, A., Sakač, M., Milovanović, I. & Sedej, I., (2011)
593 Development of a rapid resolution HPLC method for the separation and determination of 17
594 phenolic compounds in crude plant extracts, *Central European Journal of Chemistry*., 9(1) , 133-
595 142
596
597 Panusa, A, Petrucci, R., Marrosu, G., Multari, G. & Gallo, F.R., (2015) UHPLC-PDA-ESI-TOF/MS
598 metabolic profiling of *Arctostaphylos pungens* and *Arctostaphylos uva-ursi*. A comparative study of
599 phenolic compounds from leaf methanolic extracts, *Phytochemistry* 115, 79–88.
600
601 Placer, Z.A., Custman, N., & Hohnson, B.C. (1968). Estimation of product of lipid peroxidation
602 (malonyldialdehyde) in biochemical system. *Analytical Biochemistry*, 16, 359-364.
603
604 Rodrigo, R., Rivera, G., Orellana, M., Araya, J., & Bosco, C. (2002). Rat kidney antioxidant
605 response to long-term exposure to flavonol rich red wine. *Life Sciences*,71(24), 2881–2895.
606
607 Rojas-Rivera, J.,Ortiz, A., & Egidio, J. (2012). Antioxidants in Kidney Diseases: The Impact of
608 Bardoxolone Methyl. *International Journal of Nephrology*, (1-12). Article ID 321714.
609
610 Saeidi, J., Bozorgi. H., Zendehtel, A., & Mehrzad, J. (2012).Therapeutic effects of aqueous extracts
611 of *Petroselinum crispum* on ethylene glycol-induced kidney calculi in rats. *Urology Journal* 9(1),
612 361-366.
613

614 Saito, C., Zwingmann, C., & Jaeschke, H. (2010). Novel mechanisms of protection against
615 acetaminophen hepatotoxicity in mice by glutathione and N-acetylcysteine. *Hepatology*,
616 51(1),246-54.
617

618 Sarepoua, E., Tangwongchai, R., Suriharn, B., & Lertrat, K. (2013) Relationships between
619 phytochemicals and antioxidant activity in corn silk. *International Food Research Journal* 20(5),
620 2073-2079.
621

622 Sarepoua, E., Tangwongchai, R., Suriharn, B., & Lertrat, K. (2015) Influence of variety and harvest
623 maturity on phytochemical content in corn silk, *Food Chemistry* 169, 424–429.
624

625 Schinella, G., Troiani, G., Dávila, V., Buschiazzo, P.M., & Tournier, H.(2000). Antioxidant effects
626 of an aqueous extract of *Ilex paraguariensis*. *Biochemical and Biophysical Research*
627 *Communication*, 269,357–60.
628

629 Sedlak, I., & Lindsay, R.H. (1968). Estimation of total protein-bound and non protein sulfhydryl
630 groups in tissue with Elman reagent. *Analytical Biochemistry*, 16, 259-205.
631

632 Shahidi, F., (1997) Natural antioxidants: an overview, in: *Natural antioxidants*, (pp. 1-11)
633 F. Shahidi, ed., AOCS Press, Champaign, IL.
634

635 Shalaby, M.A., Abd-Elkhalik, A., & Hammouda, A.E. (2014). Nephroprotective, diuretic and
636 antioxidant effects of some medicinal herbs in gentamicin-nephrotoxic rats. *Journal of Intercultural*
637 *Ethnopharmacology*, 3(1),1-8.
638

639 Shikov, A.,N., Pozharitskaya, O.N., Makarov, V.,G., Wagner, H., Rob Verpoorte, R. & Heinrich,
640 M. (2014). Medicinal Plants of the Russian Pharmacopoeia; their history and applications. *Journal*
641 *of Ethnopharmacology*,154 (3), 481–536.
642

643 Snedecor, G.W., & Cochran, W.G. (1986). *Statistical Methods* ,The 7th Edition, (pp 90-93.) Ames:
644 Iowa State University Press, USA.
645

646 Sukandar, E.Y.,Sigit, J.I., & Adiwibow,L.F. (2013).Study of Kidney Repair Mechanisms of Corn
647 Silk (*Zea mays* L. Hair)-Binahong (*Anredera cordifolia* (Ten.) Steenis) Leaves Combination in Rat
648 Model of Kidney Failure. *International Journal of Pharmacology*, 9(1),12-23.
649

650 Štajner, D., Milic, N., Čanadanovic-Brunet, J., Kapor, A.J., Štajner ,M. & Popović B.M. (2006).
651 Exploring *Allium* species as a source of potential medicinal agents. *Phytotherapy Research*, 20,
652 581-584.
653

654 Štajner, D., Popović, B.M., Čanadanović-Brunet, J., Đilas, S., & Četković,G. (2014). Nutritive
655 composition and free radical scavenger activity of honey enriched with of *Rosa* spp. *LWT - Food*
656 *Science and Technology*, 55: 408-413. – a
657

658 Štajner, D., Popović, B.M., Čalić, D., & Štajner, M. (2014). Comparative Study of Antioxidant
659 Status in Androgenic Embryos of *Aesculus hippocastanum* and *Aesculus flava*. *Scientific World*
660 *Journal* , 2014,7 pages, doi:10.1155/2014/767392 –b.

661
662 Ueno, S., Kashimoto, T., Susa, N., Shiho, K., Seki, T., Ito, N., Takeda-Homma, S., Nishimura, Y.,
663 & Sugiyama, M. (2007). Estimation of hydroxyl radical generation by salicylate hydroxylation
664 method in kidney of mice exposed to ferric nitrilotriacetate and potassium bromate. *Free Radical*
665 *Research*,41(11),1246-1252.

666 Vinson, J.A., Proch, J., & Bose, P. (2001). Determination of quantity and quality of polyphenol
667 antioxidants in foods and beverages. *Methods in Enzymology*, 335 , 103-114.

668 Wu, Y., Tian, Q., Laihao, L., Khan , M,N., Yang, X., Zhang, Z., Hu, X., & Chen, S. (2013). Inhibitory
669 effect of antioxidant peptides derived from *Pinctada fucata* protein on ultraviolet-induced photoaging
670 in mice. *Journal of functional foods* ,5, 527–538.

671
672 Yildiz, L., Baskan, K.S., Tutem, E., Apek, R., (2008) Combined HPLC-CUPRAC (cupric ion
673 reducing antioxidant capacity) assay of parsley, celery leaves, and nettle, *Talanta* 77, 304–313.

674
675 Zhang, J., Chu, C.J., Li , X.L., Yao , S., Yan ,B., Ren , H.L. Xu ,N.Y., Liang , Z.T., & Zhao, Z.Z. ,
676 (2014), Isolation and identification of antioxidant compounds in *Vaccinium bracteatum* Thunb. by
677 UHPLC-Q-TOF LC/MS and their kidney damage protection. *Journal of Functional Foods*,1162-70.

678
679
680
681
682
683
684
685
686
687
688
689
690
691
692
693
694
695

696
697
698

Table 1. The effects of corn silk, parsley and bearberry extracts on diuresis, excreted K⁺ and Na⁺ quantities.

Day	0	1	7	14	21	28
Corn silk extract						
Diuresis (ml)	3.52 ^{ab}	4.1 ^d	3.8 ^c	3.58 ^{bc}	3.47 ^a	3.45 ^a
K ⁺ (mmol/l)	202.48 ^b	233.3 ^d	219.17 ^c	200.7 ^b	194.7 ^a	201.61 ^b
Na ⁺ (mmol/l)	317.68 ^b	334.10 ^c	318.09 ^b	312.09 ^a	310.98 ^a	316.71 ^b
Parsley extract						
Diuresis (ml)	3.59 ^b	3.98 ^c	4.18 ^d	3.49 ^a	3.5 ^a	3.57 ^b
K ⁺ (mmol/l)	196.87 ^b	239.05 ^c	218.3 ^d	192.46 ^b	184.39 ^a	203.66 ^c
Na ⁺ (mmol/l)	347.98 ^b	397.01 ^a	450.0 ^d	361.93 ^c	346.69 ^b	354.98 ^b
Bearberry extract						
Diuresis (ml)	3.32 ^b	3.41 ^c	3.35 ^b	3.25 ^a	3.28 ^a	3.30 ^{ab}
K ⁺ (mmol/l)	185.98 ^c	211.71 ^d	209.19 ^d	179.69 ^b	181.91 ^{bc}	166.79 ^a
Na ⁺ (mmol/l)	330.72 ^d	353.93 ^c	334.24 ^d	300.92 ^c	270.85 ^a	290.93 ^b
Control (water)						
Diuresis (ml)	3.48 ^{bc}	3.45 ^a	3.51 ^c	3.49 ^{ab}	3.46 ^a	3.50 ^c
K ⁺ (mmol/l)	200.86 ^c	198.49 ^{bc}	195.30 ^a	201.0 ^c	197.93 ^{ab}	200.0 ^c
Na ⁺ (mmol/l)	333.39 ^{bc}	326.0 ^a	335.73 ^c	329.19 ^{ab}	331.9 ^b	333.32 ^{bc}

702 Values were means for ten mice. Values with the same letter, in each colon, are not significantly different
703 according to Duncan test (p < 0.05).
704

705
706
707 Table 2. Effects of extracts of *Maydis stigma*, *Petroselinum crispum* and *Uvae ursi folium* on blood
708 urea nitrogen and serum creatinine contents.
709

Investigated extract	Urea (mg/l)	Creatinine(mg/l)
Corn silk	8,50 ^c	19,00 ^c
Parsley	10,40 ^b	30,00 ^b
Bearberry	11,40 ^{ab}	68,0 ^a
Control (water)	12,6 ^a	67,5 ^a

711 Values were means for ten mice. Values with the same letter, in each colon, are not significantly different
712 according to Duncan test (p < 0.05).

713
 714 Table 3. Effects of extracts of *Maydis stigma*, *Petroselinum crispum* and *Uvae ursi folium* extracts
 715 on DPPH radical scavenging capacity of mice kidneys.
 716

Extract	Kidney DPPH (IC ₅₀ ; mg/cm ³)
Control	4.64 ^a
Bearberry	3.19 ^b
Parsley	2.62 ^c
Corn silk	3.48 ^b

717 Values were means for ten mice. Statistical comparisons between samples were performed with Duncan t-
 718 test. Differences were considered significant at p < 0.05.
 719

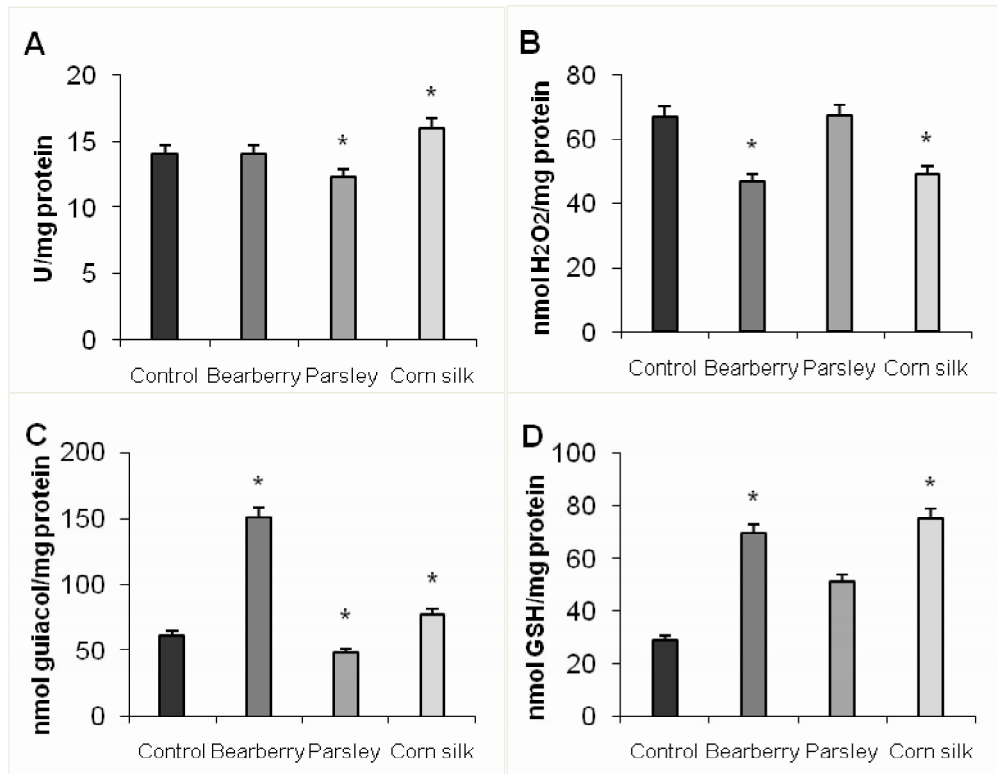
720
 721
 722 Table 4. Content of plant phenolics in crude parsley, bearberry and corn silk extracts are expressed
 723 as mg g⁻¹ extract
 724

Phenolic compound	parsley	bearberry	corn silk
gallic acid	0.16 ^a	7.75 ^b	0.15 ^a
protocatechuic acid	0.56	-	-
catechin and catechin derivates	0.16 ^a	12.23 ^b	-
caffeic acid	0.46	-	-
vanillic acid	0.55 ^a	-	0.50 ^a
chlorogenic acid	0.11	-	-
epicatechin	0.40	-	-
ferulic acid	-	-	0.32
rutin and rutin derivates		13.74	
quercetin and quercetin derivates	1.86 ^b	0.47 ^a	-
kaempferol and kaempferol derivates	5.84	0.26	-
apigenin and apigenin derivates	0.30	-	-
luteolin and luteolin derivates	-	0.23	-
myricetin and myricetin derivates	-	0.12	-
orientin derivates	-	-	3.24
total	10.39 ^b	34.80 ^c	4.21 ^a

725 All samples were extracted and analyzed in triplicate. Values with the same letter, in each row, are
 726 not significantly different according to Duncan test (p < 0.05).
 727

728
 729
 730
 731
 732
 733

734



735
736

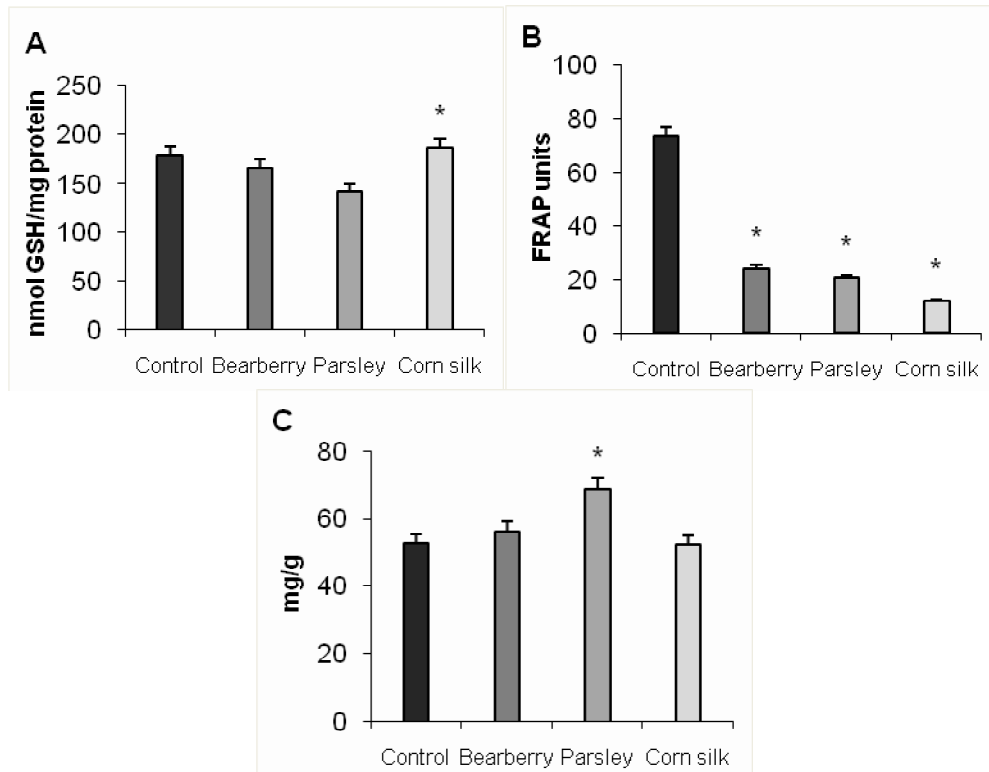
737 **Fig. 1** - Effects of extracts of *Maydis stigma*, *Petroselinum crispum* and *Uvae ursi folium* extracts,
738 on antioxidant enzymes activities in mice kidneys: A) Superoxide dismutase, SOD; B) Catalase,
739 CAT; C) Guaiacol peroxidase, GPx and D) Glutathione peroxidase, GSH-Px

740 Values were means for ten mice. Statistical comparisons between samples were performed with Duncan t-test.

741 * Marked differences were considered as significant at p < 0.05 from control group.

742
743
744
745
746
747
748
749
750
751
752
753
754
755
756
757
758
759
760
761

762



763

764

765 **Fig. 2.** Effects of extracts of *Maydis stigma*, *Petroselinum crispum* and *Uvae ursi folium* extracts on
766 glutathione quantity (A), ferric reducing antioxidant power (B) and soluble protein content (C) in
767 mice kidneys.

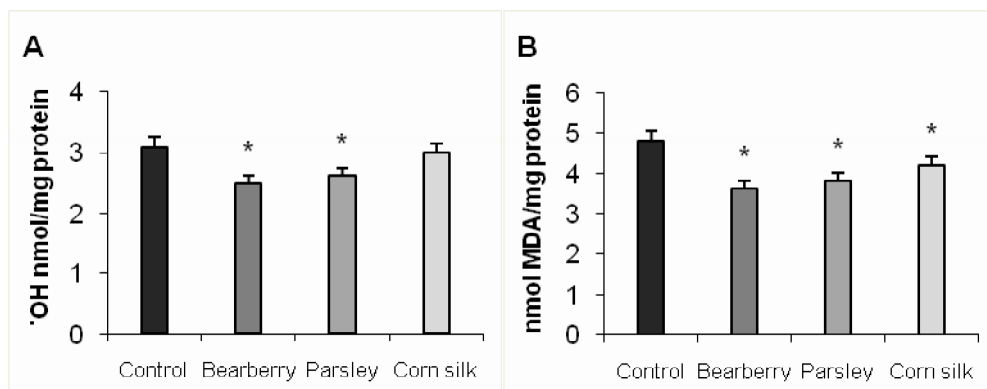
768 Values were means for ten mice. Statistical comparisons between samples were performed with Duncan t-test.

769 * Marked differences were considered as significant at $p < 0.05$ from control group.

770

771

772



773

774

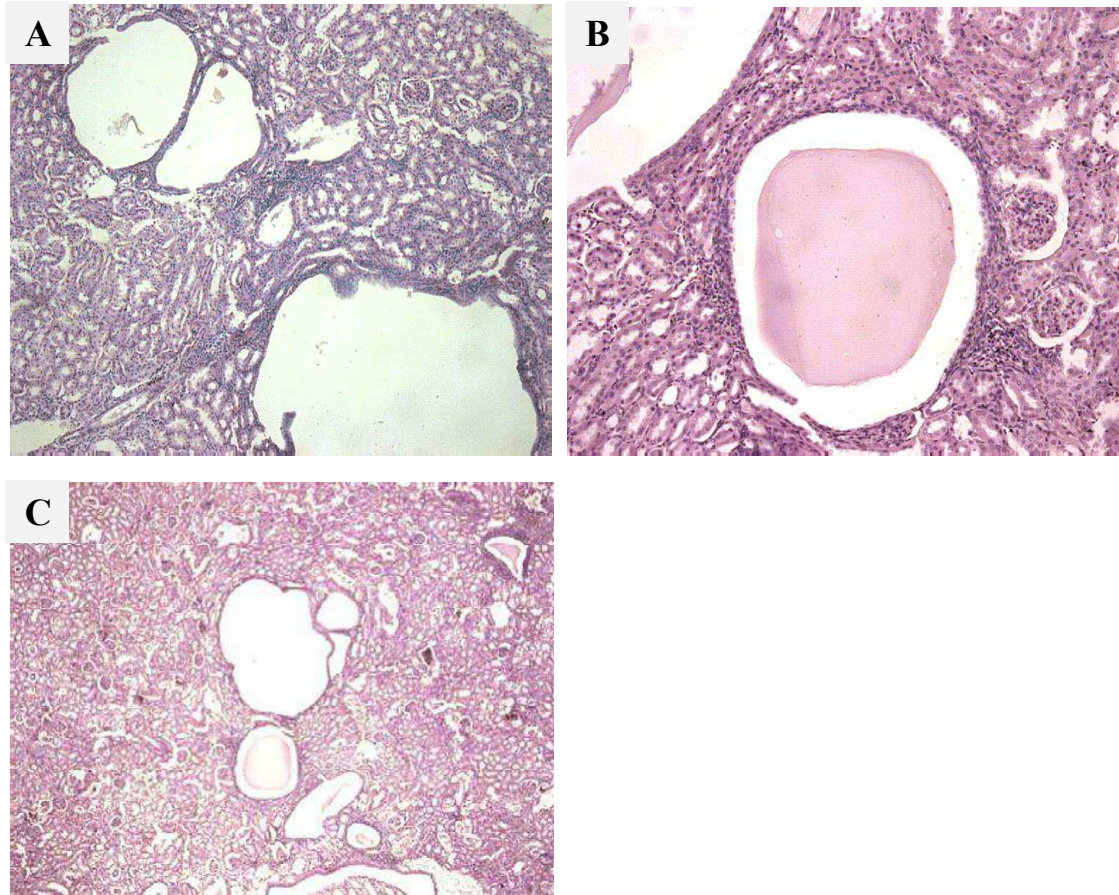
775 **Fig. 3.** Effects of extracts of *Maydis stigma*, *Petroselinum crispum* and *Uvae ursi folium* extracts on
776 hydroxyl radical quantity (A) and lipid peroxidation (B) in mice kidneys.

777 Values were means for ten mice. Statistical comparisons between samples were performed with Duncan t-test.

778 * Marked differences were considered as significant at $p < 0.05$ from control group.

779

780



781
782

783
784

785 Fig.4. Histopathological features of kidney tissue after *Uva ursi* extract administration
786 A- microcystic changes in cortical region, 100x, H&E ; B - hyalic cylinders in cystic formation
787 200x, H&E; C- microcystic changes in cortical region, 50x, H&E.