



TITLE: Prediction of the genetic similarity of wheat and wheat quality by reversed-phase High-Performance liquid Chromatography and Lab-on-Chip methods

AUTHORS: Aleksandra Torbica, Daniela Horvat, Dragan Živančev, Miona Belović, Gordana Šimić, Damir Magdić, Nevena Đukić, Krešimir Dvojković

This article is provided by author(s) and FINS Repository in accordance with publisher policies.

The correct citation is available in the FINS Repository record for this article.

NOTICE: This is the author's version of a work that was accepted for publication in *Acta Alimentaria*. Changes resulting from the publishing process, such as peer review, editing, corrections, structural formatting, and other quality control mechanisms may not be reflected in this document. Changes may have been made to this work since it was submitted for publication. A definitive version was subsequently published in *Acta Alimentaria*, Volume 46, Issue 2, June 2017, Pages 137–144. DOI: 10.1556/066.2016.0003

This item is made available to you under the Creative Commons Attribution-NonCommercial-NoDerivative Works – CC BY-NC-ND 3.0 Serbia



Prediction of the genetic similarity of wheat and wheat quality by reversed-phase High-Performance liquid Chromatography and Lab-on-Chip methods

Short running title: RP-HPLC and LoAC gluten proteins quantification

A. Torbica^{a*}, D. Horvat^b, D. Živančev^a, M. Belović^a, G. Šimić^b, D. Magdić^c, N. Đukić^d, K. Dvojković^b

^aInstitute of Food Technology, University of Novi Sad, Bulevar cara Lazara 1, 21000 Novi Sad, Serbia

^bAgricultural Institute Osijek, Južno Predgrađe 17, 31000 Osijek, Croatia

^cFaculty of Food Technology, J.J. Strossmayer University of Osijek, Franje Kuhača 18, 31000 Osijek, Croatia

^dFaculty of Natural Science, University of Kragujevac, Radoja Domanovića 12, 34000 Kragujevac, Serbia

*To whom correspondence should be addressed:

Phone: +381 21 485 3625; fax: +381 21 450 725; e-mail: aleksandra.torbica@fins.uns.ac.rs.

The aim of this study was to compare efficiency of RP-HPLC (High-Performance Liquid Chromatography) and LoAC (Lab-on-a-Chip) methods for wheat gluten protein quantification regard clustering of wheat cultivars according to the genetic similarity (HMW-GS combinations), as well as to explore relations of these two methods to wheat quality parameters. For that purpose, wheat quality parameters (protein content, falling number, wet gluten content, gluten index, Farinograph, Extensograph, and Amylograph) and amounts of gliadin and glutenin fractions by RP-HPLC and LoAC methods were determined in two different sets of wheat cultivars (Croatian and Serbian). The percentages of gluten proteins and the values of quality parameters were used to characterize the samples by principal component analysis (PCA). Gluten protein quantification performed by method based on the protein fraction separation by molecular weights (LoAC) was better in grouping of genetically similar wheat cultivars than quantification of proteins separated by their different solubility in specified solvent gradient (RP-HPLC). LoAC method showed higher potential in wheat quality prediction.

Key words: wheat quality, RP-HPLC, Loac, gluten proteins, genetic similarity

Wheat flour possesses numerous different utilizations in food industry that depend upon the type of desired product. When it is mixed with water it forms dough that possesses unique viscoelastic properties due to the formation of gluten. Gluten consists of two different types of proteins: monomeric gliadins and polymeric glutenin. These complex compounds are most responsible for the viscoelastic properties of dough and baking quality of wheat (PANOZZO et al., 2001). Gliadins are classified into α -gliadins, β -gliadins and ω -gliadins on the basis of NH₂-terminal amino acid sequences (KASARDA et al., 1983; BIETZ et al., 1977), whereas glutenins are classified into high-molecular-weight glutenin subunits (HMW-GS) and low-molecular-weight glutenin subunits (LMW-GS) on the basis their mobility on Sodium Dodecyl Sulfate-Polyacrylamide Gel Electrophoresis (SDS-PAGE) (CORNISH et al., 2006; BIETZ et al., 1975). The molecular weights of HMW-GS determined by SDS-PAGE are in the range of 80 to 130 kDa (BUNCE et al., 1985), whereas the molecular weights of LMW-GS determined by SDS-PAGE are in the range of 30 to 50 kDa (GRAS et al., 2001).

Different studies have been conducted to examine the effects of gluten proteins - glutenins (MACRITCHIE et al., 1991; PAYNE et al., 1984; PIROZI et al., 2008) and gliadins (GIL HUMANES et al., 2012; HUEBNER et al., 1997; PAYNE et al., 1984) on the viscoelastic properties of dough and technological quality of wheat. One of the first studies on this topic was conducted by PAYNE and co-workers (1984) showed that the glutenins are responsible for the elasticity of the dough. It is well-known fact that a couple of subunits HMW-GS from D1 loci 5+10 formed stronger dough than a couple HMW-GS from D1 loci 2+12, because of an extra cysteine residue on repetitive amino acid sequence of the subunit 5 (LAFIANDRA et al., 1993). Also, ANDERSON and BÉKÉS (2011) showed that the addition of individual glutenin subunits from GluD1 loci HMW-GS of x-type (2 and 5) in five different dough samples has a greater impact on the

technological dough parameters than addition of HMW-GS of γ -type (10 and 12). Furthermore, specific composition of HMW-GS in wheat cultivars is one of the most important genetic factors which influence the rheological properties of dough (PAYNE et al., 1987). It has been recently demonstrated that composition and quantity of HMW-GS could be determined by Lab-on-a-Chip (LoaC) technique (ŽIVANČEV et al., 2013; 2015), whereas for accurate determination of their amounts a combined extraction - Reversed-Phase High-Performance Liquid Chromatography (RP-RP-HPLC) procedure developed by WIESER and co-workers (1998) is still used.

The influence of LMW-GS composition on dough properties has not been studied as much as the influence of HMW-GS composition. The study of LUO and co-workers (2001) showed that the specific composition of LMW-GS has a greater impact on the dough extensibility than on the dough strength, and these properties were directly related to the HMW-GS.

It is believed that the monomeric gliadins act as plasticizers of the polymeric gluten system and thus provide plasticity/viscosity to wheat dough (KHATKAR et al., 1995). Addition of the gliadin fractions to the wheat flours significantly reduced the maximum resistance and increases the extensibility of dough (SCHROPP & WIESER, 1996). In opposite to this study research of the GIL-HUMANES and co-workers (2012) showed that the content of γ -gliadin was positively correlated with the dough development as measured by Mixolab, which contributes to the dough strength.

For the determination of dough extensibility Extensigraph (INDRANI et al., 2011; ZHANG et al., 2007) is still used, whereas for the determination dough mixing properties Farinograph still represents common rheological method (INDRANI et al., 2011; RAKSZEGI et al., 2008).

The aim of this paper was to compare efficiency of two most applied methods for relative quantification of gluten proteins regard clustering of wheat cultivars according to the genetic

similarity (HMW-GS combinations), as well as to explore relations of these two methods to wheat quality parameters.

1. Material and methods

1.1. Materials

Nine bread wheat (*Triticumaestivum*L.) cultivars (“Divana”, “Aida”, “Felix”, “Seka”, “Renata”, “Soissons”, “Olimpija”, “Vulkan”, and “Tihana”) grown in Croatia at the Agricultural Institute in Osijek and nine bread wheat cultivars (“Dragana”, “Ljiljana“, “Pobeda”, “Bastijana”, “Nevesinjka”, “Simonida”, “Etida”, “Zvezdana”, and NS3-5299/2) grown in Serbia at the Institute of Field and Vegetable Crops in Novi Sad harvested in season 2009 were investigated in the present study.

1.2. Samples preparation and analytical procedure

Protein content (P) was determined on wheat kernels by FOSS Infratec 1241 Grain Analyzer (FOSS ANALYTICAL AB, Hillerød, Denmark), whereas Falling number (FN) was determined by Falling Number 1600 (PERTEN INSTRUMENTS, Huddinge, Sweden) according to ICC standard method 107/1. Wheat samples were milled by MLU – 202 (Bühler, Uzwil, Switzerland) and obtained flours were used for further rheological analyses. Wet gluten content (WG) and gluten index (GI) were determined by Glutomatic 2100 (PERTEN INSTRUMENTS, Huddinge, Sweden) according to ICC standard method 155. Rheological quality of flour samples were determined by Farinograph, Extensograph and Amylograph (C.W. BRABENDER, Duisburg, Germany) according to (MSZ6369/6-1988, ICC114/1, ICC126/1 respectively). The extractions of gliadin and glutenin subunits for Loac and reversed-phase high-performance liquid chromatography (RP-RP-HPLC) methods as well as

LoaC and RP-RP-HPLC analyses were performed according to ŽIVANČEV and co-workers (2015).

1.3. Data analysis

The data were statistically analysed by STATISTICA 12.0 software (StatSoft Inc., USA, 2013). Descriptive statistics was used to explore the percentage amounts of gluten proteins as well as rheological parameters and for that purpose, mean values, ranges and coefficients of variation (CV) were calculated. The percentages of gluten proteins and the values of quality parameters were used to characterize the samples by principal component analysis (PCA). The PCA was performed on the symmetric correlation matrix. Pearson correlation coefficients were calculated in order to further discuss the relationships between examined variables.

2. Results and discussion

2.1 Quantification of protein fractions

The quantitative results of gluten protein fractions obtained by RP-RP-HPLC and LoaC methods were summarized in Table 1. In general, the results of gliadin subunits show much better agreement between these two examined methods than results of glutenin subunits. Also, the results obtained by RP-RP-HPLC method are less variable since SD of all protein fractions gained by RP-RP-HPLC method are lower than SD gained by LoaC method.

2.2 Technological quality of wheat

Regarding the wheat end-use quality parameters (Table 2), a large variability of some analyzed parameters was noticed. The range of WG, FN, DDT, Stab, R, DS, E and EXT varied between weak or medium to excellent, which indicate significant differences among dough rheology of examined wheat cultivars. In opposition to them the range of P, GI, WA, R_{5min} , R_{5min}/EXT , Y and SV showed lower variation.

2.3 *Wheat cultivars HMW-GS composition*

Allelic variation at *Glu-1* loci of examined wheat cultivars obtained by Loac method is shown in Table 3. The most frequent HMW-GS combinations of Croatian cultivars were 2* 7+8 5+10 and 2* 7+9 5+10 (22.22%, for both combinations), whereas the most frequent combinations HMW-GS of Serbian cultivars were 2* 7+9 5+10 and 7+9 2+12 (33.33%, for both combinations).

2.4 *Principal component analysis of wheat genetic similarity and wheat quality prediction*

For the purpose of statistical analysis, whole data set was divided into two parts; quantification results from electrophoregrams (Loac) and quantification results from chromatograms (RP-HPLC). Both data sets were correlated with technological quality parameters of wheat flour (data not shown).

Principal Component Analysis (PCA) was applied in order to reduce the initial complex data set to smaller number of independent variables which represent the linear combination of all examined quality parameters. Therefore all variables than did not contribute significantly to explanation of data set variability were excluded from further considerations.

The first two principal components (Fig. 1a – loading plot) explained 79.09 % of reduced data set total variability, which incorporated the results of gluten fractions quantification obtained by RP-HPLC method. Variation of HMW and LMW percentages (which are negatively correlated one with another) caused change of WA, WG, and GI values, as well as extensogram parameters R_{5min}/EXT and R_{5min} . On the basis of vector position it could be seen that WA was in significant positive correlation ($p < 0.05$) with percentage of HMW subunits and in significant negative correlation ($p < 0.05$) with LMW percentages. On the other hand, GI was positively correlated ($p < 0.05$) with R_{5min} , and negatively correlated ($p < 0.05$) with WG.

The score plot (Fig. 1b) showed that the cultivars were mostly grouped by WG, LMW and HMW percentages, but Croatian and Serbian assortments were not completely separated.

The first two PCs (Fig. 2a–loading plot) explained 75.58% of reduced data set total variability, which included the results of gluten fractions quantification obtained by LoAC method. In this case, variation of all gluten fractions percentages (both gliadins and glutenins) caused change in GI value and extensogram parameters E, R_{5min}/EXT and R_{5min} . The above mentioned parameters GI and E were in significant positive correlation ($p < 0.05$) with R_{5min} .

The score plot (Fig. 2b) did not show that cultivars were grouped by amount of particular protein fraction, but Croatian and Serbian assortments were completely separated. The exception was cultivar “Tihana”, which was the only Croatian cultivar with 2+12 subunit combination, possessed by majority of Serbian cultivars.

Correlation coefficients between the percentage share of protein fractions and tested wheat technological quality parameters were higher when LoAC method was applied. However, these dependencies were not statistically significant.

3. Conclusions

The aforementioned results indicated that gluten protein quantification performed by method based on the protein fraction separation by molecular weights (LoAC) was better in grouping of genetically similar wheat cultivars than quantification of proteins separated by their different solubility in specified solvent gradient (RP-HPLC). LoAC method showed higher potential in wheat quality prediction.

***Acknowledgment**

This work was supported by the Ministry of Education, Science and Technological Development of the Republic of Serbia within the Project of technological development no.

TR 31007 and bilateral cooperation between Serbia and Croatia—Genetic polymorphism of gluten proteins and its relationship to bread-making quality of wheat (*Triticum aestivum* L.). Dr. Nikola Hristov from Institute of Field and Vegetable Crops (Novi Sad).

References

- Anderson, O. D. & Békés, F. (2011): Incorporation of high-molecular-weight glutenin subunits into doughs using 2 gram mixograph and extensigraphs. *Journal of Cereal Science*, 54, 288–295.
- Bietz, J. A., Huebner, F. R., Sanderson, J. E., Wall, J. S. (1977): Wheat Gliadin Homology Revealed through N-Terminal Amino Acid Sequence Analysis. *Cereal Chemistry*, 54, 1070-1083.
- Bietz, J. A., Shepherd, K. W., Wall, J. S. (1975): Single kernel analysis of glutenin: Use in wheat genetics and breeding. *Cereal Chemistry*, 52, 513-532.
- Bunce, N. A. C., White, R. P., Shewry, P. R. (1985): Variation in Estimates of Molecular Weights of Cereal Prolamins by SDS-PAGE. *Journal of Cereal Science*, 3, 131–142.
- Cornish, G. B., Békés, F., Eagles, H. A., Payne, P. I. (2006): Gliadin and Glutenin. Wrigley, C., Bushuk, W. (Eds.), *The Unique Balance of Wheat Quality*, AACC International, St. Paul, MN, USA, pp. 243–280.
- Gil-Humanes, J., Pistón, F., Rosell, C. M., Barro, F. (2012): Significant down-regulation of γ -gliadins has minor effect on gluten and starch properties of bread wheat. *Journal of Cereal Science*, 56, 161–170.
- Gras, P. W., Anderssen, R. S., Keentock, M., Bekes, F., Appels, R. (2001): Gluten protein functionality in wheat flour processing: a review. *Australian Journal of Agricultural Research*, 52, 1311 - 1323.

- Gupta, R. B., & Shepherd, K. W. (1990): Two-step one-dimensional SDS-PAGE analysis of LMW subunits of glutelin. 1. Variation and genetic control of the subunits in hexaploid wheats. *Theoretical and Applied Genetics*, 80, 65–74.
- Huebner, F. R., Nelsen, T. C., Chung, O. K., Bietz J. A. (1997): Protein Distributions Among Hard Red Winter Wheat Varieties as Related to Environment and Baking Quality. *Cereal Chemistry*, 74, 123-128.
- ICC standard method 107/1. Determination of the "Falling Number" according to Hagberg
- ICC standard method 155. Determination of Wet Gluten Quantity and Quality (Gluten Index ac. to Perten) of Whole Wheat Meal and Wheat Flour (*Triticumaestivum*)
- ICC Standard No. 114/1, Method for using the BrabenderExtensograph
- ICC Standard No. 126/1, Method for using the BrabenderAmylograph
- Indrani, D., Swetha, P., Soumya, C., Rajiv, J., Venkateswara Rao, G. (2011): Effect of multigrains on rheological, microstructural and quality characteristics of north Indian parotta – An Indian flat bread. *LWT – Food Science and Technology*, 44, 719–724.
- Kasarda, D. D., Autran, J.-C., Lew, E. J.-L., Nimmo, C. C., Shewry, P. R. (1983): N-terminal amino acid sequences of w-gliadins and w-secalins: implications for the evolution of prolamin genes. *BiochimicaetBiophysicaActa*, 747, 138-150.
- Khatkar, B. S., Bell, A. E., Schofield, J. D. (1995): The dynamic rheological properties of gluten and gluten sub-fractions from wheats of good and poor bread making quality. *Journal of Cereal Science*, 22, 29–44.
- Lafiandra, D., D'Ovidio, R., Porceddu, E., Margiotta, B., Colaprico, G. (1993): New data supporting high molecular glutenin subunit 5 as the determinant of quality differences among the pairs 5+10 vs. 2+12. *Journal of Cereal Science*, 18, 197-205.

- Luo, C., Griffin, W. B., Branlard, G., McNeil, D. L. (2001): Comparison of low- and high molecular-weight wheat glutenin allele effects on flour quality. *Theoretical and Applied Genetics*, 102, 1088–1098.
- MacRitchie, F., Kasarda, D. D., Kuzmick, D. D. (1991): Characterization of wheat protein fractions differing in contributions to breadmaking quality. *Cereal Chemistry*, 68, 122–130.
- Metakovsky, E. V., Wrigley, C. W., Bekes, F., Gupta, R. B. (1990): Gluten polypeptides as useful genetic markers of dough quality in Australian wheats. *Australian Journal of Agricultural Research*, 41, 289–306.
- MSZ 6369/6-1988. Flour testing methods. Determination of water absorption capacity and baking quality, Hungary.
- Panozzo, J. F., Eagles, H. A., Wootton, M. (2001): Changes in protein composition during grain development in wheat. *Australian Journal of Agricultural Research*, 52, 485–493.
- Payne, P. I., Holt, L. M., Jackson, E. A., Law, C. N., Damania, A. B. (1984): Wheat Storage Proteins: Their Genetics and Their Potential for Manipulation by Plant Breeding [and Discussion]. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*, 304, 359–371.
- Payne, P. I., Nightingale, M. A., Krattiger, A. F., Holt, L. M. (1987): The relationship between HMW glutenin subunit composition and the bread-making quality of British-grown wheat varieties. *Journal of the Science of Food and Agriculture*, 40, 51–65.
- Pirozi, M. R., Margiotta, B., Lafiandra, D., MacRitchie, F. (2008): Composition of polymeric proteins and bread-making quality of wheat lines with allelic HMW-GS differing in number of cysteines. *Journal of Cereal Science*, 48, 117–122.

- Rakszegi, M., Pastori, G., Jones, H. D., Békés, F., Butow, B., Láng, L., Bedő, Z., Shewry, P. R. (2008): Technological quality of field grown transgenic lines of commercial wheat cultivars expressing the 1Ax1 HMW glutenin subunit gene. *Journal of Cereal Science*, 47, 310–321.
- Schropp, P., & Wieser, H. (1996): Effects of high molecular weight subunits of glutenin on the rheological properties of wheat gluten. *Cereal Chemistry*, 73, 410–413.
- Shewry, P. R., Halford, N. G., Tatham, A. S. (1992): *Journal of Cereal Science*, 15, 105–120.
- Wieser, H., Antes, S., Seilmeier, W. (1998): Quantitative determination of gluten protein types in wheat flour by reversed-phase high-performance liquid chromatography. *Cereal Chemistry*, 75, 644–650.
- Wrigley, C. W., & Beitz, J. A. (1988): Proteins and amino acids. Pomeranz, Y. (Ed.), *Wheat Chemistry and Technology* (3rd ed.), AACC International, St. Paul, MN, USA, pp. 159–275.
- Zhang, P., He, Z., Chen, D., Zhang, Y., Larroque, O. R., Xia, X. (2007): Contribution of common wheat protein fractions to dough properties and quality of northern-style Chinese steamed bread. *Journal of Cereal Science*, 46, 1–10.
- Živančev, D. R., Nikolovski, B. G., Torbica, A. M., Mastilović, J. S., Đukić, N. H. (2013): Lab-on-a-chip method uncertainties in determination of high-molecular-weight glutenin subunits. *Chemical Industry and Chemical Engineering Quarterly*, 19, 553–561, 2013.
- Živančev, D., Horvat, D., Torbica, A., Belović, M., Šimić, G., Magdić, D., Đukić, N. (2015): Benefits and Limitations of Lab-on-a-Chip Method over Reversed-Phase High-Performance Liquid Chromatography Method in Gluten Proteins Evaluation. *Journal of Chemistry*, vol. 2015, Article ID 430328.

Table 1. Quantification of protein fractions performed by RP-RP-HPLC and LoaC methods

Protein fractions	RP-RP-HPLC			LoaC		
	Mean	Range	SD	Mean	Range	SD
% $\alpha + \gamma$ subunit	7.32	4.40–15.73	2.31	7.34	3.10–17.32	3.59
% ω subunit	92.72	84.26–95.60	2.31	92.55	82.68–96.90	3.61
% HMW-GS	27.60	20.75–37.50	3.52	13.26	4.66–29.57	5.50
% LMW-GS	72.39	62.43–79.25	3.52	86.84	70.43–95.34	5.49
% HMW/LMW	38.45	26.18–60.07	6.97	15.75	4.89–41.98	7.83

Table 2. Wheat end-use quality parameters

Parameter	Mean	Range	SD
P (% d.w.)	13.6	12.1–16.4	0.9
WG (%)	29.7	22.7–40.0	4.6
GI (%)	96.4	81.0–99.8	5.0
FN (s)	302.9	111.0–436.0	74.0
WA (%)	61.9	56.9–67.0	3.0
DDT (min)	3.1	1.5–10.3	2.0
Stab (min)	2.3	0.1–7.0	2.2
R (min)	6.5	1.6–30.0	6.5
DS (FU)	55.5	2.4–110.0	29.3
E (cm²)	85.8	47.0–128.0	27.3
R_{5min} (EU)	235.8	140.0–350.0	54.1
EXT (mm)	170.7	132.0–209.0	21.2
R_{5min}/EXT	1.4	0.7–2.1	0.4
Y (%)	72.1	67.0–76.0	2.2
SV (cm³/g)	3.4	3.0–3.8	0.2

List of abbreviations: P (% d.w.) – protein content per dry weight, WG (%) – wet gluten content, GI – gluten index, FN (s) – falling number, WA (%) – water absorption, DDT (min) – dough development time, Stab (min) – dough stability, R (min) – dough resistance, DS (FU) – degree of softening, E (cm²) – dough energy, R_{5min} (EU) – dough resistance on 5 minutes, EXT (mm) – dough extensibility, R_{5min}/EXT – ratio of dough resistance on 5 minute and extensibility, Y (%) – flour extraction yield, SV (cm³/g) – specific volume of bread.

Table 3. Composition of HMW-GS in wheat cultivars

Cultivar	GLU-A1	GLU-B1	GLU-D1
Soissons	2*	7 + 8	5 + 10
Felix	2*	7 + 8	5 + 10
Divana	2*	7 + 9	5 + 10
Zlata	2*	7 + 9	5 + 10
Seka	1	7 + 9	5 + 10
Vulkan	N	7 + 8	5 + 10
Renata	1	7 + 8	5 + 10
Aida	2*	17 + 18	5 + 10
Tihana	1	7 + 9	2 + 12
Etida	N	7 + 9	5 + 10
Ljiljana	N	7 + 9	5 + 10
Pobeda	2*	7 + 9	5 + 10
Bastijana	2*	7 + 9	5 + 10
NS3-5299	2*	7 + 9	5 + 10
Simonida	N	7 + 9	2 + 12
Dragana	N	7 + 9	2 + 12
Zvezdana	N	7 + 9	2 + 12
Nevesinjka	2*	7 + 8	5 + 10 & 2 + 12

Figure captions

Figure 1. PCA plot of relationship between the glutenin subunits' percents determined by RP-HPLC method and quality parameters (a) and differentiation between the cultivars (b) after the reduction of variables

Figure 2. PCA plot of relationship between the glutenin subunits' percents determined by LoaC method and quality parameters (a) and differentiation between the cultivars (b) after the reduction of variables

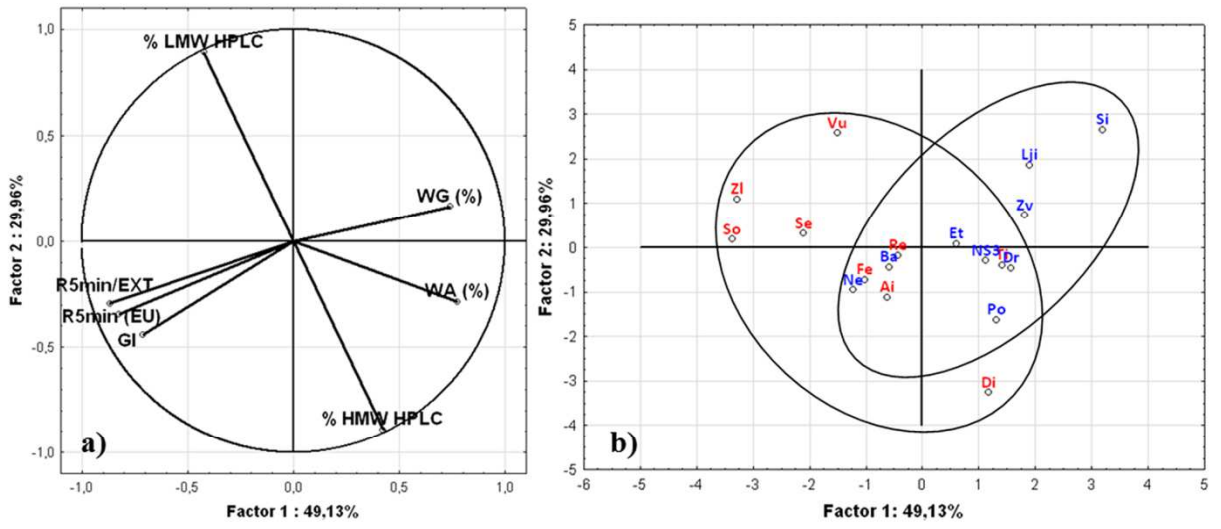


Figure 1

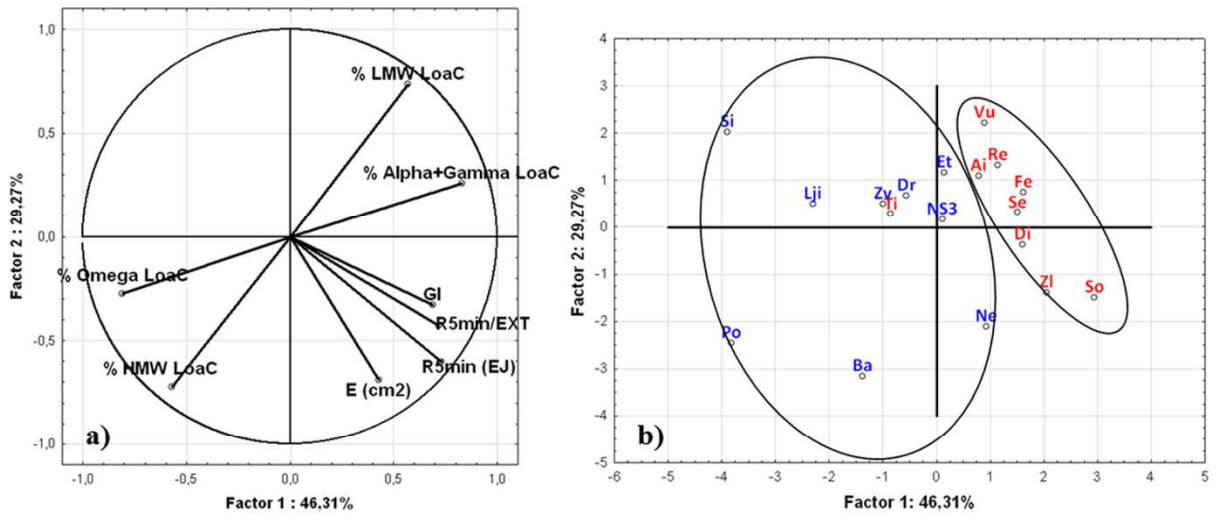


Figure 2